# 1,4-Naphthalenediyl-Bridged Molecular Gyrotops: Rotation of the Rotor and Fluorescence in Solution

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**S** Supporting Information

[AB](#page-7-0)STRACT: [Macrocage m](#page-7-0)olecules with a bridged  $\pi$ -electron system have been reported as molecular gyrotops in which the  $\pi$ -electron system can rotate within the cage. We recently reported the dynamics of the rotor in solution using 1,4 naphthalenediyl-bridged molecular gyrotops, which consist of cages formed of three  $C_{14}$ ,  $C_{16}$ , or  $C_{18}$  chains. In this work, we synthesized novel gyrotops with  $C_{15}$  and  $C_{17}$  chains and systematically investigated the activation energies for the



rotation of the rotor in solution. The activation energies for rotation in solution were found to decrease with increasing size of the cage. Therefore, a rotational barrier can be designed by adjusting the length of the side chains in these molecular gyrotops. Additionally, these gyrotops were fluorescent in solution; the quantum yields and lifetimes of the fluorescence were investigated. However, these properties were not influenced by the chain length owing to a large difference in time scale between fluorescence  $(10^{-8}-10^{-9} \text{ s})$  and the rotational dynamics inside the cage  $(10^{\circ}-10^{-5} \text{ s})$ .

## **ENTRODUCTION**

Artificial molecular rotors that show rotation of a part of the molecule have recently become a widely investigated topic because the molecular motion could affect the properties and reactivity of these molecules. The chemical field of artificial molecular machines aims to design novel molecular-based materials in which these motions are controlled.<sup>1−7</sup> For this reason, macrocage molecules with a bridged  $\pi$ -electron system have been reported as molecular gyroscopes a[nd](#page-7-0) gyrotops because the  $\pi$ -electron system can rotate rapidly inside a cage framework.<sup>2−5</sup> The synthesis, structure, and dynamics of molecular gyroscopes and gyrotops have been investigated by Garcia-Gar[ibay](#page-7-0) et al.,  $2.6$  Gladysz et al., 3 and our group.<sup>4</sup>

We synthesized a crystalline molecular gyrotop 1 with a phenylene rotor and [sh](#page-7-0)owed the rota[tio](#page-7-0)n of the rotor [ins](#page-7-0)ide a cage, accompanied by a thermal change in the birefringence of the single crystal.<sup>4e</sup> The effects of the chain length  $(C_{14}, C_{16})$ and  $C_{18}$  chains) on the dynamic properties of the molecular gyrotops in a cr[yst](#page-7-0)alline state have recently been reported.<sup>4a</sup> Phenylene is suitable for the gyrotop rotor in a crystalline state because it is a symmetric and small  $\pi$ -electron syste[m.](#page-7-0) However, it is difficult to investigate the dynamics of the rotor in solution due to the axial symmetry of the rotor.

We recently reported the synthesis and structure of 1,4 naphthalenediyl-bridged molecular gyrotops (C14, C16, and C18 in Figure 1) and revealed the chain length  $(C_{14}, C_{16},$  and  $C_{18}$  chains) dependence of the rotation of the rotor in  $solution.<sup>4d</sup>$  As the 1,4-naphthalenediyl rotor is axially asymmetric, slow rotation of the rotor inside the cage, at less than the [NM](#page-7-0)R time scale, can be easily observed as asymmetric



Figure 1. (a) A model gyrotop. (b) Structural formula of molecular gyrotops 1 and C14−C18.

NMR signals from the cage framework. In contrast, rapid rotation of the rotor is confirmed by symmetric NMR signals from the cage due to chemical exchange within the asymmetric structure.

In this report, novel molecular gyrotops (C15 and C17 in Figure 1) with medium alkyl chains  $(C_{15}$  and  $C_{17}$  chains) forming the cages were designed and synthesized. Synthesis of the cage revealed that the yield had a remarkable chain length dependence. The activation energies for rotation of the rotor were estimated by investigating the temperature dependence of the line shape of the <sup>13</sup>C NMR signal and <sup>13</sup>C<sup>−13</sup>C exchange spectroscopy (EXSY) NMR. $8,9$  Thus, the effects of cage size on the barrier to rotation of the naphthalenediyl rotor were revealed systematically a[nd](#page-7-0) accurately. Additionally, the fluorescence properties, such as the quantum yields and lifetimes, of these gyrotops in solution were investigated.

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However, these properties were found to be unaffected by the chain length owing to a large difference in time scale between fluorescence and the rotational dynamics inside the cage. The basic studies of the properties of these simple caged compounds will allow implementation of novel principles for the molecular design of new functional molecules. For this reason, a series of cage-like cyclophanes containing a fluorophore were synthesized, and their basic characteristics as functional molecules were investigated.

## ■ RESULTS AND DISCUSSION

Synthesis of Molecular Gyrotops. Synthesis of 1,4naphthalenediyl-bridged molecular gyrotops (C14, C16, and C18) with even-numbered alkyl chain lengths  $(C_{14}, C_{16}, and)$  $C_{18}$  chains) forming the cage was previously reported (Scheme 1, route  $A$ ).<sup>4d</sup> They were synthesized by ring-closing metathesis<sup>10</sup> (RCM) reaction of bissilylnaphthalene (6–8) and subsequent [hyd](#page-7-0)rogenation of the unsaturated cage. The cages were [sep](#page-7-0)arated from the mixture by preparative gel permeation chromatography (GPC). The structural identification of the cages was reported in the previous report. $4d$  The structure of C14i was identified from  $^{13}$ C NMR spectra. Owing to the structural symmetry of C14i, 14 meth[yle](#page-7-0)ne signals were observed, half of which had signal intensities twice as large as the other half. The  $^{13}$ C NMR spectra of C16i and C18i showed similar signal patterns, although some of the signals were overlapped. As these isomers (C14i, C16i, and C18i) were obtained as oils, structural analysis by X-ray crystallography could not be carried out. For reference, the crystal structures of the phenylene derivatives of C18i were reported previously.<sup>4b</sup>

To synthesize molecular gyrotops with odd-numbered alkyl chain lengths (C15 and C17), introduction of differ[ent](#page-7-0) trialkenylsilyl groups to naphthalene is necessary. Therefore, stepwise silylations of naphthalene were performed (Scheme 1, route B). Monolithiation of 1,4-dibromonaphthalene with n-BuLi, followed by a substitution reaction with tri(8-nonenyl) chlorosilane (4), afforded 1-bromo-4-silylnaphthalene 9 in a 47% yield. Bissilylnaphthalenes (10 and 11) were obtained from 9 in 53% and 54% yields, respectively, via similar lithiation and substitution reactions. The RCM reaction of 10 and 11 in the presence of Grubbs' first-generation catalyst, followed by hydrogenation of the unsaturated cage, gave the corresponding desired cages [C15 (14% yield) and C17 (13% yield)] along with the formation of structural isomers [C15i (29% yield) and C17i (28% yield)]. The cages and isomers were separated by preparative gel permeation chromatography (GPC). The structures of the cages were identified by NMR spectroscopy and single-crystal X-ray diffraction studies, as described later. The structural identification of the isomers (C15i and C17i) was difficult because the NMR signals of the methylenes were intricately overlapped owing to the different lengths of the two methylene chains adjacent to the same silicon in C15i ( $C_{14}$  and  $C_{16}$  chains) and C17i ( $C_{16}$  and  $C_{18}$  chains). Hence, the structures were assigned using the MS spectra and in comparison with the NMR spectra of C14i.

The product ratios and yields of the cages are summarized in Table 1. A remarkable chain length dependence of the product

Table 1. Yields of the Cages and Their Isomers in RCM Reactions of Bissilylnaphthalenes (6, 7, 8, 10, and 11)

	yield $(\%)$		$ratio^{\alpha}$		
precursor	cage	isomer	cage:isomer		
6 $(n = 14)^b$	29	27	5:5		
10 $(n = 15)$	14	29	3:7		
7 $(n = 16)^{b}$		32	2:9		
11 $(n = 17)$	13	28	3:7		
8 $(n = 18)^{b}$	18	15	5:5		
<sup><i>a</i></sup> Product ratio. <sup><i>b</i></sup> Reference 4d.					

ratios in the RCM r[eac](#page-7-0)tion of bissilylnaphthalenes was observed. The cage/isomer product ratios are larger than the statistic ratios in the synthesis of C14, C15, C17, and C18; this is because the statistical ratio of formation of the cage and the isomer, derived from probability theory, in this RCM reaction was cage/isomer =  $1/3$ .<sup>4b</sup> The reason for the predominant formation of the isomer in the synthesis of C16 remains unclear. It is possible tha[t t](#page-7-0)he odd length of the alkenyl chains  $(C_9$  chains) in the precursor may prevent formation of the cage. A similar chain length dependence of the cage/isomer product ratio was observed in the synthesis of the phenylene-bridged derivatives.<sup>4</sup>

Molecular Structure Determined by Single-Crystal Xray Diffr[ac](#page-7-0)tion Studies. Single-crystal X-ray diffraction studies were performed to determine the molecular structure of the crystalline state. Recrystallization to obtain a single

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Figure 2. Molecular structures of molecular gyrotops C14−C18 as determined by X-ray crystallography (ball-and-stick model). Hydrogen atoms are omitted for clarity. Molecular structure (top row) and crystal structure (bottom row): (a)  $C14, ^{4d}$  (b)  $C15$ , (c)  $C16, ^{4d}$  (d)  $C17,$  (e)  $C18. ^{4d}$ 

crystal of the cages (C14−C18) was conducted by evaporation from a tetrahydrofuran (THF)/methanol  $(4:1 \text{ v/v})$  solution. Unfortunately, the isomers (C14i−C18i) were not crystallized owing to their oily nature. We previously reported the molecular structures of C14, C16, and C18 as determined by X-ray crystallography.<sup>4d</sup> In this study, the structures of C15 and C17 were analyzed; however, the structures were not refined well (i.e., final R ind[ice](#page-7-0)s of greater than 10%), because of the following undesirable characteristics. A rotational twin existed in the single crystal of C15, of which the crystal system and space group are monoclinic and  $P2<sub>1</sub>/c$ , respectively, because the  $\beta$  angle of the unit cell is close to 90°. For C17, intense reflection data could not be collected even at low temperature (90 K) because the single crystals were too thin.

The molecular structures of molecular gyrotops C14−C18 as determined by X-ray crystallography are shown in Figure 2. In all the compounds, the bissilylnaphthalene moiety was effectively surrounded by three alkyl chains. Comparing the molecular structures among molecular gyrotops C14−C18, we observed that the shape of the cage was deformed from the spherical shape in the compounds containing long alkyl chains (e.g., C18). The calculated densities of the crystals decreased with expansion of the cage size:  $1.047$  g/cm<sup>3</sup> for C14,  $1.040$  g/ cm<sup>3</sup> for C15, 1.026 g/cm<sup>3</sup> for C16 [solvent molecules (THF) were included in this crystal],  $1.027$  g/cm<sup>3</sup> for C17 [the observed temperature (90 K) differed from the others (120 K)], and 1.019  $g/cm<sup>3</sup>$  for C18, most likely because the void spaces between the cage and the rotor increased with increasing cage size.

All of the crystal structures of the molecular gyrotops (C14− C18) have similar features, such that the molecules could be stacked along their rotation axes. As described in the previous report, π-stacked aggregates of molecules were formed in the crystal structure of C16 and C18.<sup>4d</sup> However, the other molecules (C14, C15, and C17) existed as a single molecule in the crystal. The melting points of the [pre](#page-7-0)sent molecules (C14− C18) were nearly identical, although their crystal structures differ slightly.

Rotation of a 1,4-Naphthalenediyl Rotor in Solution. The rotation of the 1,4-naphthalenediyl rotor of the molecular gyrotops in solution was investigated by temperature-dependent NMR spectroscopy. Although chemically inequivalent methylene signals of the molecular gyrotops overlapped in the <sup>1</sup>H NMR spectra (see the Supporting Information), the

corresponding m[eth](#page-7-0)ylene signals of t[he](#page-7-0)  $^{13}$ C NMR spectr[a w](#page-7-0)ere well-resolved (Figure 3). Hence, the rotation of the rotor was



Figure 3.  $^{13}$ C NMR spectra (125 MHz) of compounds (a)  $C14, ^{4d}$  (b) C15, (c) C16,<sup>4d</sup> (d) C17, and (e) C18<sup>4d</sup> in toluene- $d_8$  at 300 K (methylene region). The signals of methylene carbons adjac[ent](#page-7-0) to silicon atoms a[re](#page-7-0) indicated by the symbol  $\alpha$ .

studied using 13C NMR spectroscopy. The alkyl region (10−35 ppm) of the  $^{13}$ C NMR spectra of the molecular gyrotops, recorded at 300 K in toluene- $d_8$ , is shown in Figure 3. The signals of methylenes adjacent to silicon, which are indicated by the symbol  $\alpha$  in Figure 3, were assigned by analyses of the H<sup>-13</sup>C HSQC and <sup>1</sup>H<sup>-13</sup>C HMBC NMR spectra. For instance, the signal assignments in C15 were carried out as follows: In the HSQC spectrum (Figure S18), the carbons corresponding to the signals at 13.0 and 15.3 ppm are connected to the protons correspon[ding to the](http://pubs.acs.org/doi/suppl/10.1021/acs.joc.5b01489/suppl_file/jo5b01489_si_001.pdf) signals at 1.03 and 1.09 + 0.79 (magnetically inequivalent protons) ppm, respectively. In the HMBC spectrum (Figure S19), only these

methylene protons show cross-peaks with the naphthalene carbons connected to silicon ( $\delta$  136.9). These observations indicate that the methylenes corresponding to the signals at 13.0 and 15.3 ppm are located closest to the naphthalene ring.

In the  $^{13}$ C NMR spectrum of C14 (Figure 3, a), 14 methylene signals were observed, half of which had twice the intensity of the other half.<sup>4d</sup> This indicates th[at two of t](#page-2-0)he three alkyl chains are identical on the time scale of the NMR [e](#page-7-0)xperiment and that the cage of C14 possesses quasi- $C_{2v}$ symmetry. Hence, the rotation of the naphthalene ring is very slow, or stopped, inside the cage. In the  $^{13}C$  NMR spectrum of C15 (Figure 3, b), the observed spectral features were identical to those of C14, indicating slow dynamics of the rotor. In the spec[trum of](#page-2-0)  $C16$  (Figure 3, c), the methylene signals were broadened owing to the restricted rotation of the rotor.<sup>4d</sup> In the spectrum of C18 ([Figure 3,](#page-2-0) e), nine methylene signals were observed, indicating that the three alkyl chains were [ide](#page-7-0)ntical on the NMR time [scale owi](#page-2-0)ng to rapid rotation of the internal naphthalenediyl rotor inside the cage, as shown in a previous report.<sup>4d</sup> Similar spectral features were observed in the spectrum of  $C17$  (Figure 3, d), indicating rapid rotation of the rotor.

To estimate the energy barrier for the rotation of the rotor, temperature-dependent <sup>13</sup>C [NMR](#page-2-0) spectra were investigated. As described in the previous report,  $^{13}$ C NMR spectra of C16 in toluene- $d_8$  showed a remarkable temperature dependence of the line shapes of the methylene signals.<sup>4d</sup> Simulation of the spectral line shapes assuming a three-site exchange model (Figure 4) allowed estimation of the exc[ha](#page-7-0)nge rate constants



Figure 4. Schematic representation of the three-site exchange model of the dynamics of the molecular gyrotop C16.

 $(k_{ex})$  at each observed temperature. The thermodynamic parameters for the exchange for C16 were estimated to be  $\Delta H^{\ddagger} = 9.42 \pm 0.08$  kcal mol<sup>-1</sup> and  $\Delta S^{\ddagger} = -12.0 \pm 0.3$  cal mol<sup>-1</sup> K<sup>-1</sup> by using an Eyring plot.<sup>4d</sup>

Temperature-dependent  $^{13}$ C NMR spectra of C17 in toluene- $d_8$  were investigated (Fig[ure](#page-7-0) 5). The line shapes of the methylene signals exhibited a remarkable temperature dependence. An inverse gate pulse sequence was used for the experiments to suppress the nuclear Overhauser effect in the spectra. Two signals at 12.2 and 15.1 ppm assignable to methylenes adjacent to silicon atoms, i.e.,  $\alpha$ -methylenes in Figure 3d, at 210 K showed remarkable temperature dependence. With increasing temperature, these signals broadened and [coalesced](#page-2-0) at around 230 K and became a sharp singlet above ambient temperature. The exchange rate constants  $(k_{ex})$  at each temperature were estimated by line shape analysis assuming the three-site exchange model shown in Figure 4. The activation



Figure 5. (a) Temperature-dependent  $^{13}$ C NMR spectra of C17 in toluene- $d_8$  ( $\alpha$ -methylene). Left: observed spectra. Right: simulated spectra with designated exchange rate constants  $(k_{ex})$ . (b) Eyring plot for naphthalenediyl rotation in C17. The  $k_{ex}$  values are determined from temperature-dependent <sup>13</sup>C NMR spectroscopy.

enthalpy and entropy for the rotation of the rotor in C17 were estimated to be  $\Delta H^{\ddagger}$  = 7.49 ± 0.26 kcal mol<sup>-1</sup> and  $\Delta S^{\ddagger}$  =  $-11.0 \pm 1.0$  cal mol<sup>-1</sup> K<sup>-1</sup>, respectively, from the linear plot of  $\ln(k_{ex}/T)$  versus  $1/T$  (Eyring plot, Figure 5b).

The temperature dependence of the  $^{13}$ C NMR spectra (1Dcomplete) of C15 was also investigated; however, the line shapes of the methylene signals remained unchanged. Hence, the very slow chemical exchange processes in C15 were investigated by 13C−13C EXSY NMR. EXSY NMR is useful for analysis of slow chemical exchanges, which are observed as intensity differences between exchangeable signals of crosspeaks and diagonal peaks.<sup>8,9</sup> Figure 6a shows the temperaturedependent <sup>13</sup>C<sup>−13</sup>C EXSY NMR spectra of C15 in toluene- $d_8$ with a mixing time [of](#page-7-0) [400 ms.](#page-4-0) Cross-peaks between exchangeable methylene protons adjacent to silicon atoms, i.e.,  $\alpha$ -methylenes in Figure 3b, were observed, and the intensities of the signals were remarkably temperature-dependent. Although the excha[nge proce](#page-2-0)ss is strictly represented by a three-site exchange mechanism (Figure 4), it reduces to a twosite exchange (eq 1), as two of the three structures were indistinguishable. Figure 6 also shows the rate constants  $(k_{ex})$ for the chemical exchange (eq 1). The estimated exchange constants, which [are comp](#page-4-0)arable to the exchange rates for the three-site exchange model (Figure 4), can be estimated by analyzing the signal intensities of the cross-peaks and diagonal peaks using the EXSY Calc software.<sup>11</sup> The activation enthalpy  $(\Delta H^{\ddagger})$  and entropy  $(\Delta S^{\ddagger})$  for the rotation of the rotor in C15 were estimated to be 14.6  $\pm$  2.78 k[cal](#page-7-0) mol<sup>-1</sup> and -11.9  $\pm$  8.7 cal mol<sup>-1</sup> K<sup>-1</sup>, respectively, from the Eyring plot (Figure 6b).

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Figure 6. Temperature-dependent 13C−13C EXSY NMR spectra of C15 in toluene- $d_8$  ( $\alpha$ -methylenes) with a mixing time of 400 ms. Exchange rate constants  $(k_{ex})$  estimated by EXSY Calc software are also indicated. (b) Eyring plot for naphthalenediyl rotation in C15.



The dynamics of the naphthalenediyl rotor in C14 is expected to be slower than that of C15. Therefore, the <sup>13</sup>C−<sup>13</sup>C EXSY NMR spectrum of C14 was also investigated. However, no cross-peaks between exchangeable methylene protons were observed (Figure S44). Hence, the dynamics is considered to stop or be much slower than the NMR time scale.

Unfortu[nately, the](http://pubs.acs.org/doi/suppl/10.1021/acs.joc.5b01489/suppl_file/jo5b01489_si_001.pdf) energy barriers for the rotation of the rotor of C14 and C18 could not be estimated. The temperature dependence of the  $^{13}$ C NMR spectra of C14 and C18 were reinvestigated, and the results are shown in Figures S41 and S42 and Table S2 in the Supporting Information. The chemical shifts of the signals were slightly changed by varying the temperature, indicating [that the equilibrium c](http://pubs.acs.org/doi/suppl/10.1021/acs.joc.5b01489/suppl_file/jo5b01489_si_001.pdf)age structures were temperature-dependent. However, the details of the structural change were not apparent. The signal line widths of the methylene signals, as well as the aromatic signals, were broadened with decreasing temperature. The line broadening is not ascribed to rotational dynamics of the rotor, but to the increasing viscosity of the solvent.

The estimated activation enthalpies  $(\Delta H^{\ddagger})$  and entropies  $(\Delta S^{\ddagger})$  for the rotation of the 1,4-naphthalenediyl rotor in the molecular gyrotops are summarized in Table 2. It is apparent

Table 2. Thermodynamic Parameters for Rotor Rotation in Molecular Gyrotops C14−C18

	rotation parameters <sup>b</sup>		
gyrotop	$\Delta H^{\ddagger}/\text{kcal}$ mol <sup>-1</sup>	$\Delta S^{\ddagger}/\text{cal}$ mol <sup>-1</sup> K <sup>-1</sup>	$k_{\rm av}/s^{-1}$ at 300 K
$C14^a$	$>14.6^\circ$	с	< 0.37
C15	$14.6 + 2.78$	$-11.9 + 8.7$	0.37
$C16^a$	$9.42 \pm 0.08$	$-12.0 \pm 0.3$	2200
C17	$7.46 \pm 0.10$	$-11.2 \pm 0.4$	60 000
C18 <sup>a</sup>	$< 7.46$ <sup>c</sup>	c	>60000
	$\mathbf{L}$		

 ${}^a$ Reference 4d.  ${}^b$ The errors are standard deviations.  ${}^c$ The parameter could not be determined uniquely.

that the a[cti](#page-7-0)vation enthalpies for rotation in the molecular gyrotops (14.6 ± 2.78 kcal mol<sup>-1</sup> for C15, 9.42 ± 0.08 kcal mol<sup>-1</sup> for C16, and 7.46  $\pm$  0.10 kcal mol<sup>-1</sup> for C17) decreased with increasing length of the alkyl side chains of the cage. The large negative  $\Delta S^{\ddagger}$  values (−11.9  $\pm$  8.7 cal mol<sup>-1</sup> K<sup>-1</sup> for C15,  $-12.0 \pm 0.3$  cal mol<sup>-1</sup> K<sup>-1</sup> for C16, and  $-11.2 \pm 0.4$  cal mol<sup>-1</sup>  $K^{-1}$  for C17) for rotation of the naphthalene ring inside the cage indicate a tight dynamic transition state.

Fluorescence Properties of the Cage and Noncage Isomers in Solution. Naphthalene is known as one of the fluorophores having  $\pi$ -electron systems. The fluorescence properties of silylnaphthalenes in solution have been reported so far. $^{12}$  In this study, the effects of the cage on the fluorescence properties, i.e., the quantum yield and florescence lifetime, of 1,4-bi[ssi](#page-7-0)lylnaphthalenes were investigated using the molecular gyrotops (C14−C18) and their isomers (C14i−C18i).

Figure 7 shows absorption spectra of a hexane solution of bissilylnaphthalenes between 230 and 360 nm. An intense



Figure 7. Absorption spectra of 1,4-bissilylnaphthalenes in n-hexane (230−360 nm) [np (black broken), TMSnp (black solid), C14 (purple solid), C14i (purple broken), C15 (blue solid), C15i (blue broken), C16 (green solid), C16i (green broken), C17 (orange solid), C17i (orange broken), C18 (red solid), C18i (red broken)].

absorption band around 290 nm with vibronic structures is assignable to Platt's  $L_a$  band, and a weak band at 317 nm is assignable to the  $L_b$  band. The absorption bands of caged naphthalenes (C14−C18) and their isomers (C14i−C18i) were slightly red-shifted compared with those of 1,4-bis- (trimethylsilyl)naphthalene (TMSnp).

Figure 8 shows the fluorescence spectra of the bissilylnaphthalenes in degassed hexane irradiated at 265 nm. Fluorescence b[ands with](#page-5-0) vibronic structures from localized excited states were observed. The fluorescence band maxima for all the bissilylnaphthalenes were 332 nm. The fluorescence quantum

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Figure 8. Fluorescence spectra of 1,4-bissilylnaphthalenes in  $n$ -hexane (degassed 1.0  $\times$  10<sup>-6</sup> M solution, irradiated at 265 nm) [np (black broken), TMSnp (black solid), C14 (purple solid), C14i (purple broken), C15 (blue solid), C15i (blue broken), C16 (green solid), C16i (green broken), C17 (orange solid), C17i (orange broken), C18 (red solid), C18i (red broken)].

yields ( $\Phi$ ) and fluorescence lifetimes ( $\tau_0$ ) of the bissilylnaphthalenes in hexane solution (10  $\mu$ M) were also investigated. To suppress fluorescence quenching by oxygen completely, the samples were degassed by a freeze−pump−thaw method before the measurements. The results are summarized in Table 3. The

Table 3. Fluorescence Parameters of Bissilylnaphthalenes C14−C18

compound	$\lambda_{\rm max}^{\rm a}/\rm nm$	$\Phi^b$	$\tau_0^{\rm d}/\text{ns}$
np	323	0.10 <sup>c</sup>	$115.6 \pm 8.5$
TMSnp	331	0.11	$36.7 + 2.2$
C14	332	0.12	$23.5 \pm 2.2$
C14i	332	0.11	$26.5 \pm 2.4$
C15	332	0.12	$23.3 \pm 2.2$
C15i	332	0.12	$27.0 + 2.0$
C <sub>16</sub>	332	0.10	$26.7 + 2.3$
C <sub>16i</sub>	332	0.12	$27.0 \pm 2.1$
C17	332	0.10	$25.1 \pm 2.0$
C17i	332	0.11	$26.3 \pm 2.3$
C18	332	0.11	$26.1 \pm 2.7$
C18i	332	0.11	$26.1 \pm 2.0$

a Quantum yields were determined by comparing fluorescence intensity of naphthalene in hexane ( $\Phi$  = 0.10, ref 13) under irradiation by 262 nm light. <sup>b</sup>Reference 13. <sup>c</sup>Lifetimes were measured under irradiation by 280 nm light with a time-corr[ela](#page-7-0)ted single-photon counting apparatus.

quantum yields of the cages (C14−C18) and the isomers (C14i−C18i) were almost identical within a range of 0.10− 0.12, and the fluorescence lifetimes of the cages and isomers were also identical within a range of 23.5−27.0 ns. The results indicate that the fluorescence properties, such as the band maxima, quantum yields, and lifetimes, of the cage and noncage isomers showed no dependence on the side chain length. Because the time scale of the fluorescence  $(10^{-8}-10^{-9} s)$  is much faster than that of the dynamics of the naphthalene rotor  $(10^{\circ}-10^{-5} s)$ , as described above, these fluorescence properties of the caged naphthalenes were not influenced by the dynamics of the naphthalenediyl rotors inside the cage.

#### ■ CONCLUSIONS

We systematically investigated the chain length  $(C_{14}, C_{15}, C_{16})$  $C_{17}$ , and  $C_{18}$  chains) dependence of the rotation of the rotor in solution using 1,4-naphthalenediyl-bridged molecular gyrotops. Specifically, molecular gyrotops with odd-length alkyl chains (C15 and C17) were synthesized by stepwise introduction of different trialkenylsilanes into naphthalene, followed by RCM and hydrogenation reactions. The structures of the molecular gyrotops were confirmed by X-ray crystallography, and the calculated densities of the crystals were found to decrease with increasing cage size, most likely owing to expansion of the void spaces inside the cage. The rotation of the naphthalenediyl rotor inside the cage in solution was investigated by temperature-dependent 13C NMR spectroscopy. The activation energies for the rotation in solution were found to decrease with increasing cage size. The fluorescence properties, such as the band maxima, quantum yields, and fluorescence lifetimes, of the cages as well as the noncage isomers were also investigated. However, these properties were not influenced by the side chain length because the time scale of the fluorescence  $(10^{-8} 10^{-9}$  s) is much faster than that of the dynamics of the naphthalene rotor (10°−10<sup>−</sup><sup>5</sup> s). Although it is considerable that collision between the alkyl chains consisted of the cage and the naphthalene fluorophore quenches the fluorescence of the naphthalene, the quantum yields of the fluorescence in gyrotops were not depend on the size of the chains. The observation of no cage dependence on the fluorescence properties of these molecules is important for their development as functional molecules.

#### **EXPERIMENTAL SECTION**

General. All reactions were conducted under an argon atmosphere. The chemical shifts of  ${}^{1}H$  and  ${}^{13}C$  NMR spectra are based on the residual solvent resonances, except for  $29\text{Si}$  NMR, in which the chemical shifts were referenced to external tetramethylsilane. 13C−13C EXSY NMR spectra were recorded using a NOESY pulse sequence with a mixing time of 400 ms and initial delay of 8 s. HRMS analyses were carried out by electrospray ionization (ESI) using an FT-ICR-MS system. The sample solutions for HRMS were prepared in a mixture of chloroform/methanol with a small amount of NaI. Preparatory GPC was carried out using a recycling preparative chromatograph; chloroform stabilized with ethanol was used as the eluent. Fluorescence spectra were obtained for samples that were degassed by argon bubbling before the measurements. Fluorescence quantum yields, lifetime measurements, and quenching experiments were carried out with samples thoroughly degassed by several freeze− pump−thaw sequences.

Materials. Commercially available reagents were used without further purification. Grubbs' first-generation catalyst, which is commercially available, was used for all the RCM reactions because, under reflux in dichloromethane, this catalyst was determined to be optimal for the synthesis of the phenylene-bridged derivatives previously reported in ref 4b. Trialkenylchlorosilanes (3, 4, and 5), bissilylnaphthalenes  $(6, 7,$  and  $8)$ , and molecular gyrotops  $(C14, C16,$ and C18) were synthesized according to literature procedures.<sup>4</sup>

Synthesis of 1-Bromo[-4-](#page-7-0)(tri-8-nonenylsilyl)naphthalene (9). p-Dibromonaphthalene (1.00 g, 3.50 mmol) and dry diethylet[he](#page-7-0)r (90 mL) were placed in a Schlenk flask (200 mL). n-BuLi solution (1.6 M in hexane, 2.0 mL, 1.0 equiv) was added dropwise to the flask at −78 °C. The reaction mixture turned yellow and was stirred for an additional 30 min, after which tri-8-nonenylchlorosilane (4, 2.00 g, 4.55 mmol) was added at −78 °C. The reaction mixture was warmed to room temperature for 1 h, and after stirring for 12 h, the mixture was hydrolyzed with dilute HCl (aq) solution and extracted with hexane. The organic layer was washed with saturated  $NAHCO<sub>3</sub>$  (aq) solution and dried over anhydrous  $Na<sub>2</sub>SO<sub>4</sub>$ . Silica gel column chromatography of the concentrated residue afforded 9 as a colorless oil (0.90 g, 1.47 mmol, 47% yield). 9: <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz)  $\delta$ 0.95−1.08 (m, 6H), 1.20−1.45 (m, 30H), 2.04 (q, J = 6.7 Hz, 6H), 4.95 (d,  $J = 10.0, 3H$ ), 5.01 (d,  $J = 17.0$  Hz, 3H), 5.81 (ddt,  $J = 17.0$ , 10.0, 6.7 Hz), 7.49 (d, J = 7.25 Hz, 1H), 7.56 (t, J = 8.5 Hz, 1H), 7.60  $(t, J = 8.5 \text{ Hz}, 1\text{H}), 7.77 \text{ (d, } J = 7.25 \text{ Hz}, 1\text{H}), 8.12 \text{ (d, } J = 8.5 \text{ Hz}, 1\text{H}),$ 

8.35 (d, J = 8.5 Hz, 1H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 125 MHz)  $\delta$  13.5 (1C), 24.0 (1C), 29.0 (1C), 29.1(2C), 33.6 (1C), 33.9 (1C), 114.2 (1C), 125.6 (1C), 126.4 (1C), 126.7 (1C), 128.2 (1C), 128.3 (1C), 129.4 (1C), 131.9 (1C), 134.4 (1C), 136.5 (1C), 138.8 (1C), 139.2 (1C); <sup>29</sup>Si NMR (CDCl<sub>3</sub>, 99.4 MHz)  $\delta$  –0.4; HRMS (ESI positive)  $m/z$ calcd for  $C_{37}H_{57}BrSi + Na^+$ : 631.33051, found: 631.33055.

Synthesis of 1-(Tri-7-octenylsilyl)-4-(tri-8-nonenylsilyl) naphthalene (10). Bissilylnaphthalene 10 was synthesized by the same procedure as 9, i.e., lithiation and silylation. In this case, the following reagents were used: 1-bromo-4-(tri-8-nonenylsilyl) naphthalene (9, 1.74 g, 2.86 mmol), THF (15 mL), n-BuLi solution (1.6 M in hexane, 2 mL, 1.0 equiv), and tri-7-octenylchlorosilane (3, 1.34 g, 3.37 mmol). Pure compound 10 (1.36 g, 1.52 mmol) was obtained as a colorless oil in 53% yield after GPC purification of the crude products. 10: a colorless oil; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz)  $\delta$ 0.91−1.08 (m, 12H), 1.15−1.45 (m, 54H), 2.01 (q, J = 7.1 Hz, 12H), 4.90−4.95 (m, 6H), 4.95−5.02 (m, 6H), 5.79 (ddt, J = 17.0, 10.0, 6.5 Hz, 3H), 5.80 (ddt, J = 17.0, 10.0, 6.5 Hz, 3H), 7.47 (dd, J = 6.3, 3.3 Hz, 2H), 7.60 (s, 2H), 8.11 (dd,  $J = 6.3$ , 3.3 Hz, 2H); <sup>13</sup>C NMR  $(CDCl_3, 125 MHz)$  δ 13.6 (1C), 24.0 (2C), 28.7 (1C), 29.0 (4C), 29.1 (1C), 33.6 (1C), 33.7 (1C), 33.8 (1C), 33.9 (1C), 114.1 (2C), 124.7 (2C), 129.0 (2C), 133.4 (2C), 137.4 (3C), 137.5 (1C), 139.2 (2C); <sup>29</sup>Si NMR (CDCl<sub>3</sub>, 99.4 MHz)  $\delta$  -1.0, -1.1; HRMS (ESI positive)  $m/z$  calcd for  $C_{61}H_{102}Si_2 + Na^+$ : 913.74122, found: 913.74123.

Synthesis of 1-(Tri-9-decenylsilyl)-4-(tri-8-nonenylsilyl) naphthalene (11). Bissilylnaphthalene 11 was synthesized by the same procedure as 10. In this case, the following reagents were used: 1-bromo-4-(tri-8-nonenylsilyl)naphthalene (9, 1.63 g, 2.79 mmol), THF (15 mL), n-BuLi solution (1.6 M in hexane, 2 mL, 1.0 equiv), and tri-9-decenylchlorosilane (5, 1.70 g, 3.43 mmol). Pure compound 11 (1.47 g, 1.50 mmol) was obtained as a colorless oil in 54% yield after GPC purification of the crude products. 11: a colorless oil;  $^{1}$ H NMR (CDCl<sub>3</sub>, 500 MHz) δ 0.90−1.05 (m, 12H), 1.15−1.45 (m, 66H), 2.02 (q, J = 7.0 Hz, 12H), 4.92 (d, J = 10.0, 6H), 4.98 (d, J = 17.0 Hz, 6H), 5.78 (ddt, J = 17.0, 10.0, 7.0 Hz, 3H), 5.79 (ddt, J = 17.0, 10.0, 7.0 Hz, 3H), 7.46 (dd, J = 6.1, 3.4 Hz, 2H), 7.59 (s, 2H), 8.10 (dd, J = 6.1, 3.4 Hz, 2H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 125 MHz)  $\delta$  13.6 (2C), 24.0 (2C), 29.0 (3C), 29.1 (2C), 29.2, 29.4, 33.7 (2C), 33.8 (2C), 114.1 (2C), 124.7 (2C), 129.0 (2C), 133.4 (2C), 137.4 (2C), 137.5 (2C), 139.2 (2C); <sup>29</sup>Si NMR (CDCl<sub>3</sub>, 99.4 MHz)  $\delta$  -1.1; HRMS (ESI positive)  $m/z$  calcd for  $C_{67}H_{114}Si_2 + Na^{\dagger}$ : 997.83513, found: 997.83522.

Synthesis of Molecular Gyrotop C15 and Its Isomer C15i. To a solution of first-generation Grubbs' catalyst (0.05 g, 0.06 mmol) in dichloromethane (600 mL) was added 1-(tri-7-octenylsilyl)-4-(tri-8 nonenylsilyl)naphthalene (10) (960 mg, 1.07 mmol) in dichloromethane (200 mL) dropwise while stirring over 12 h, and the mixture was further stirred for 12 h. The volatile materials were removed in vacuo, and the metal catalyst was removed from the toluene-soluble fraction by flash column chromatography (silica gel, benzene). Hydrogen gas (3 atm) was then introduced in an autoclave to a solution of toluene (5 mL) and the reaction mixture in the presence of 10% Pd/C (0.03 g), and the mixture was allowed to stand for 72 h at 60 °C. After the excess  $H_2$  gas was released, the mixture was filtered to remove the Pd/C. The volatile materials were removed in vacuo. The fractions containing C15 and C15i were collected separately by GPC (chloroform), and the solvents were evaporated. Pure compound C15 (120 mg, 0.15 mmol, 14% yield) was obtained as colorless crystals by recrystallization from THF/methanol (4:1). Pure compound C15i (251 mg, 0.30 mmol, 29% yield) was obtained without further purification. C15: colorless crystals; mp 100.1-101.3 °C; <sup>1</sup>H NMR  $(CDCl<sub>3</sub>$ , 500 MHz)  $\delta$  0.80−1.21 (m, 36H), 1.22−1.60 (m, 54H), 7.46 (dd,  $J = 8.0$ , 4.0 Hz, 2H, 6- and 7-positions of Naphthalene), 7.62 (s, 2H, 2- and 3-positions of Naphthalene), 8.12 (dd,  $J = 8.0$ , 4.0 Hz, 2H, 5- and 8-positions of Naphthalene); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 125 MHz)  $\delta$ 13.0 (2C), 15.3 (4C), 23.3(2C), 23.4 (4C), 27.4 (2C), 27.5 (2C), 27.6 (4C), 28.3 (6C), 28.5 (4C), 28.7 (2C), 28.8 (4C), 28.9 (1C), 29.3 (2C), 32.6 (4C), 32.7 (2C), 124.6 (2C, 6- and 7-positions of Naphthalene), 128.9 (2C, 5- and 8-positions of Naphthalene), 133.6

(2C, 2- and 3-positions of Naphthalene), 136.9 (2C, 1- and 4-positions of Naphthalene), 137.6 (2C, 4a- and 8a-positions of Naphthalene); <sup>29</sup>Si NMR (CDCl<sub>3</sub>, 99.4 MHz)  $\delta$  –0.5; HRMS (ESI positive)  $m/z$ calcd for  $C_{55}H_{96}Si_2 + Na^+$ : 835.69428, found: 835.69422. C15i: a colorless oil; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz)  $\delta$  0.90–1.25 (m, 36H), 1.25−1.60 (m, 54H), 7.50 (dd, J = 6.0, 3.0 Hz, 2H), 7.64 (s, 2H), 8.15 (dd, J = 6.0, 3.0 Hz, 2H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 125 MHz)  $\delta$  13.1, 13.3, 13.9, 14.0, 22.6, 23.3, 23.4, 26.4, 26.8, 26.9, 27.0, 27.1, 27.3, 27.6, 27.7, 28.0, 28.4, 28.5, 28.7, 27.0, 31.8, 32.7, 32.8, 32.8, 124.7, 129.0, 133.4, 137.3, 137.4; <sup>29</sup>Si NMR (CDCl<sub>3</sub>, 99.4 MHz) δ −0.7 (1Si), −0.8 (1Si); HRMS (ESI positive)  $m/z$  calcd for  $C_{55}H_{96}Si_2 + Na^+$ : 835.69428 found:835.69430.

Synthesis of Molecular Gyrotop C17. Molecular gyrotop C17 and its isomer C17i were synthesized from 11 (1.17 g, 1.20 mmol) by the same procedure as C15. Yields of the compounds were 13% for C17 (135 mg, 0.15 mmol), and 28% for C17i (296 mg, 0.33 mmol). C17: colorless crystals; mp 135.6–137.8 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz) δ 0.90−1.05 (m, 12H), 1.05−1.50 (m, 90H), 7.49 (dd, J = 6.0, 3.5 Hz, 2H), 7.66 (s, 2H), 8.13 (dd, J = 6.0, 3.5 Hz, 2H); 13C NMR (CDCl<sub>3</sub>, 125 MHz)  $\delta$  14.1 (6C), 23.7 (6C), 28.4 (18C), 28.5 (6C), 29.0 (6C), 29.2 (3C), 33.3 (6C), 124.68, 129.68, 133.66, 137.29, 137.61; <sup>29</sup>Si NMR (CDCl<sub>3</sub>, 99.4 MHz) δ −0.930; HRMS (ESI positive)  $m/z$  calcd for  $C_{61}H_{108}Si_2 + Na^{\dagger}$ : 919.78818, found: 919.78821. C17i: a colorless oil; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz)  $\delta$ 0.95−1.60 (m, 102H), 7.48−7.58 (m, 2H), 7.67 (s, 2H), 8.12−8.23 (m, 2H); 13C NMR (CDCl3, 125 MHz) δ 13.3, 13.4, 14.2, 23.3, 23.6, 23.8, 27.1, 27.4, 27.6, 27.7, 28.0, 28.1, 28.2, 28.3, 28.5, 28.8, 28.9, 29.0, 32.7, 33.3, 33.5, 124.7, 129.0, 133.5, 137.4, 137.5; <sup>29</sup>Si NMR (CDCl<sub>3</sub>, 99.4 MHz)  $\delta$  -0.9 (2Si); HRMS (ESI positive)  $m/z$  calcd for  $C_{61}H_{108}Si_2 + Na^+$ : 919.78818, found: 919.78819.

Synthesis of Molecular Gyrotop C14. Molecular gyrotop C14<sup>4d</sup> and its isomer C14i were synthesized from 6 (589 mg, 0.77 mmol) by the same procedure as C15. Yields of the compounds were 29% f[or](#page-7-0)  $C14^{4d}$  (164 mg, 0.22 mmol), and 27% for C14i (154 mg, 0.21 mmol). **C14i**: a colorless oil; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  0.80–1.21 (m, 36[H\),](#page-7-0) 1.22−1.60 (m, 48H), 7.46 (dd, J = 8.0, 4.0 Hz, 2H), 7.62 (s, 2H), 8.12 (dd, J = 8.0, 4.0 Hz, 2H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$ 13.1 (4C), 13.8 (2C), 22.6 (4C), 23.2 (2C), 26.3 (4C), 26.8 (4C), 26.9 (4C), 27.0 (4C), 27.4 (2C), 27.6 (2C), 28.1 (2C), 28.3 (2C), 31.8 (4C), 32.5 (2C), 124.7 (2C), 128.9 (2C), 133.4 (2C), 137.3 (2C), 137.6 (2C); <sup>29</sup>Si NMR (CDCl<sub>3</sub>, 79.5 MHz)  $\delta$  –0.5; Anal. Calcd for  $C_{52}H_{90}Si_2$ : C, 80.96; H, 11.76. Found: C, 80.97; H, 11.85.

**Synthesis of Molecular Gyrotop C16.** Molecular gyrotop  $C16^{4d}$ and its isomer C16i were synthesized from 7 (1.00 g, 1.07 mmol) by the same procedure as C15. Yields of the compounds were 7% f[or](#page-7-0)  $C16<sup>4d</sup>$  (63 mg, 0.073 mmol), and 32% for C16i (288 mg, 0.34 mmol). **C16i**: a colorless oil; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  0.95–1.30, 1.30– 1.68[, 7](#page-7-0).52 (dd, J = 8.0, 4.3 Hz, 2H), 7.68 (s, 2H), 8.18 (dd, J = 8.0, 4.3 Hz, 2H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  13.1, 14.1, 23.3, 23.9, 27.2, 27.6, 27.7, 28.1, 28.3, 28.4, 28.7, 32.8, 33.3, 124.7, 129.0, 133.5, 137.4; <sup>29</sup>Si NMR (CDCl<sub>3</sub>, 79.5 MHz)  $\delta$  –0.7; Anal. Calcd for C<sub>58</sub>H<sub>102</sub>Si<sub>2</sub>: C, 81.42; H, 12.02. Found: C, 81.61; H, 11.80.

Synthesis of Molecular Gyrotop C18. Molecular gyrotop  $C18<sup>4d</sup>$ and its isomer C18i were synthesized from 8 (555 g, 0.596 mmol) by the same procedure as C15. Yields of the compounds were 18% f[or](#page-7-0)  $C18^{4d}$  (101 mg, 0.11 mmol), and 15% for C18i (86 mg, 0.092 mmol). **C18i**: a colorless oil; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  0.95–1.50 (m, 108[H\)](#page-7-0), 7.49 (dd,  $J = 8.0$ , 4.3 Hz, 2H), 7.64 (s, 2H), 8.14 (dd,  $J = 8.0$ , 4.3 Hz, 2H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  13.3, 14.3, 23.6, 24.0, 27.4, 27.6, 28.1, 28.2, 28.3, 28.4, 28.5, 28.7, 28.8, 33.3, 33.6, 124.7, 129.1, 133.4, 137.4, 137.5; <sup>29</sup>Si NMR (CDCl<sub>3</sub>, 79.5 MHz) δ -0.879; Anal. Calcd for C<sub>64</sub>H<sub>114</sub>Si<sub>2</sub>: C, 81.80; H, 12.23. Found: C, 81.68; H, 12.47.

X-ray Crystallographic Analyses of Two Molecular Gyrotops. The diffraction data of molecular gyrotops C15 and C17 were collected using graphite-monochromatized Mo K $\alpha$  radiation ( $\lambda$  = 0.71069 Å). Crystallographic data for C15 (120 K): monoclinic,  $P2_1/c$ ,  $a = 22.779(6)$  Å,  $b = 12.654(3)$  Å,  $c = 18.025(5)$  Å,  $\beta = 90.008(4)$ °, V  $= 5195(2)$  Å3, R1 = 0.1058  $(I > 2\sigma I)$ , wR2 = 0.2935 (all data). Crystallographic data for C17 (90 K): triclinic,  $P\overline{1}$ ,  $a = 13.028(9)$  Å, b

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= 18.089(13) Å,  $c = 25.708(18)$  Å,  $\alpha = 74.702(12)^\circ$ ,  $\beta = 83.371(15)^\circ$ ,  $\gamma = 89.177(15)^\circ$ ,  $V = 5804(7)$  Å<sup>3</sup>, R1 = 0.1874 (I > 2 $\sigma$ I), wR2 = 0.5740 (all data). The crystallographic data were deposited at the Cambridge Crystallographic Database Centre (CCDC-1407304 for C15, and CCDC-1407258 for C17).

## ■ ASSOCIATED CONTENT

#### **6** Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.joc.5b01489.

Copies of NMR spectra for all new compounds (9, 10, 11, [C14i](http://pubs.acs.org), C15, C15i, C16i, C17, C17i[, and](http://pubs.acs.org/doi/abs/10.1021/acs.joc.5b01489) C18i). Details of temperature-dependent 13C NMR studies of C14, C15, C17, and C18. Details of fluorescence lifetime measurements of silylnaphthalenes (PDF) Crystallographic data for C15 (CIF) Crystallographic data for C17 (CIF[\)](http://pubs.acs.org/doi/suppl/10.1021/acs.joc.5b01489/suppl_file/jo5b01489_si_001.pdf)

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#### Notes

The auth[ors declare no com](mailto:wsetaka@tmu.ac.jp)peting financial interest.

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#### ■ REFERENCES

(1) (a) Vogelsberg, C. S.; Garcia-Garibay, M. A. Chem. Soc. Rev. 2012, 41, 1892. (b) Karim, A. R.; Linden, A.; Baldridge, K. K.; Siegel, J. S. Chem. Sci. 2010, 1, 102. (c) Blanzani, V.; Credi, A.; Venturi, M. Molecular Devices and Machines, 2nd ed.; Wiley-VCH: Weinheim, 2008. (d) Kay, E. R.; Leigh, D. A.; Zerbetto, F. Angew. Chem., Int. Ed. 2007, 46, 72. (e) Browne, W. R.; Feringa, B. L. Nat. Nanotechnol. 2006, 1, 25. (f) Kelly, T. R., Ed. Molecular Machines; Topics in Current Chemistry; Springer: Hidelberg, 2005; Vol. 262. (g) Kottas, G. S.; Clarke, L. I.; Horinek, D.; Michl, J. Chem. Rev. 2005, 105, 1281. (h) Garcia-Garibay, M. A. Proc. Natl. Acad. Sci. U.S.A. 2005, 102, 10771. (i) Balzani, V.; Credi, A.; Raymo, F. M.; Stoddart, J. F. Angew. Chem., Int. Ed. 2000, 39, 3348.

(2) (a) Commins, P.; Garcia-Garibay, M. A. J. Org. Chem. 2014, 79, 1611. (b) Commins, P.; Nuñ ez, J. E.; Garcia-Garibay, M. A. J. Org. Chem. 2011, 76, 8355. (c) Nuñ ez, J. E.; Natarajan, A.; Khan, S. I.; Garcia-Garibay, M. A. Org. Lett. 2007, 9, 3559.

(3) (a) Nawara-Hultzsch, A. J.; Stollenz, M.; Barbasiewicz, M.; Szafert, S.; Lis, T.; Hampel, F.; Bhuvanesh, N.; Gladysz, J. A. Chem. Eur. J. 2014, 20, 4617. (b) Zeits, P. D.; Rachiero, G. P.; Hampel, F.; Reibenspies, J. H.; Gladysz, J. A. Organometallics 2012, 31, 2854. (c) Skopek, K.; Gladysz, J. A. J. Organomet. Chem. 2008, 693, 857. (d) Skopek, K.; Barbasiewicz, M.; Hampel, F.; Gladysz, J. A. Inorg. Chem. 2008, 47, 3474. (e) Han, J.; Deng, C.; Fang, R.; Zhao, D.; Wang, L.; Gladysz, J. A. Organometallics 2010, 29, 3231. (f) Hess, G. D.; Hampel, F.; Gladysz, J. A. Organometallics 2007, 26, 5129. (g) Wang, L.; Shima, T.; Hampel, F.; Gladysz, J. A. Chem. Commun. 2006, 4075. (h) Wang, L.; Hampel, F.; Gladysz, J. A. Angew. Chem., Int. Ed. 2006, 45, 4372. (i) Nawara, A. J.; Shima, T.; Hampel, F.; Gladysz, J. A. J. Am. Chem. Soc. 2006, 128, 4962. (j) Shima, T.; Hampel, F.; Gladysz, J. A. Angew. Chem., Int. Ed. 2004, 43, 5537.

(4) (a) Setaka, W.; Inoue, K.; Higa, S.; Yoshigai, S.; Kono, H.; Yamaguchi, K. J. Org. Chem. 2014, 79, 8288. (b) Setaka, W.; Higa, S.;

Yamaguchi, K. Org. Biomol. Chem. 2014, 12, 3354. (c) Setaka, W.; Yamaguchi, K. J. Am. Chem. Soc. 2013, 135, 14560. (d) Setaka, W.; Koyama, A.; Yamaguchi, K. Org. Lett. 2013, 15, 5092. (e) Setaka, W.; Yamaguchi, K. Proc. Natl. Acad. Sci. U.S.A. 2012, 109, 9271. (f) Setaka, W.; Yamaguchi, K. J. Am. Chem. Soc. 2012, 134, 12458. (g) Shionari, H.; Inagaki, Y.; Yamaguchi, K.; Setaka, W. Org. Biomol. Chem., in press (DOI:201510.1039/C5OB01644D).

(5) (a) Setaka, W.; Ohmizu, S.; Kira, M. Chem. Commun. 2014, 50, 1098. (b) Marahatta, A. B.; Kanno, M.; Hoki, K.; Setaka, W.; Irle, S.; Kono, H. J. [Phys. Chem. C](http://dx.doi.org/10.1039/C5OB01644D) 2012, 116, 24845. (c) Setaka, W.; Ohmizu, S.; Kira, M. Chem. Lett. 2010, 39, 468. (d) Setaka, W.; Ohmizu, S.; Kabuto, C.; Kira, M. Chem. Lett. 2007, 36, 1076.

(6) (a) Dominguez, Z.; Dang, H.; Strouse, M. J.; Garcia-Garibay, M. A. J. Am. Chem. Soc. 2002, 124, 2398. (b) Godinez, C. E.; Zepeda, G.; Garcia-Garibay, M. A. J. Am. Chem. Soc. 2002, 124, 4701. (c) Dominguez, Z.; Dang, H.; Strouse, M. J.; Garcia-Garibay, M. A. J. Am. Chem. Soc. 2002, 124, 7719. (d) Dominguez, Z.; Khuong, T.-A. V.; Dang, H.; Sanrame, C. N.; Nuñ ez, J. E.; Garcia-Garibay, M. A. J. Am. Chem. Soc. 2003, 125, 8827. (e) Horansky, R. D.; Clarke, L. I.; Winston, E. B.; Price, J. C.; Karlen, S. D.; Jarowski, P. D.; Santillan, R.; Garcia-Garibay, M. A. Phys. Rev. B: Condens. Matter Mater. Phys. 2006, 74, 054306. (f) Horansky, R. D.; Clarke, L. I.; Price, J. C.; Khuong, T.- A. V.; Jarowski, P. D.; Garcia-Garibay, M. A. Phys. Rev. B: Condens. Matter Mater. Phys. 2005, 72, 014302. (g) Vogelsberg, C. S.; Bracco, S.; Beretta, M.; Comotti, A.; Sozzani, P.; Garcia-Garibay, M. A. J. Phys. Chem. B 2012, 116, 1623. (h) Czajkowska-Szczykowska, D.; Rodríguez-Molina, B.; Magaña-Vergara, N. E.; Santillan, R.; Morzycki, J. W.; Garcia-Garibay, M. A. J. Org. Chem. 2012, 77, 9970. (i) Rodríguez-Molina, B.; Pérez-Estrada, S.; Garcia-Garibay, M. A. J. Am. Chem. Soc. 2013, 135, 10388. (j) Rodríguez-Molina, B.; Ochoa, M. E.; Romero, M.; Khan, S. I.; Farfán, N.; Santillan, R.; Garcia-Garibay, M. A. Cryst. Growth Des. 2013, 13, 5107. (k) Torres-Huerta, A.; Rodríguez-Molina, B.; Hö pfl, H.; Garcia-Garibay, M. A. Organometallics 2014, 33, 354. (l) Jiang, X.; Rodríguez-Molina, B.; Nazarian, N.; Garcia-Garibay, M. A. J. Am. Chem. Soc. 2014, 136, 8871. (7) (a) Bastien, G.; Lemouchi, C.; Wzietek, P.; Simonov, S.; Zorina, L.; Rodríguez-Fortea, A.; Canadell, E.; Batail, P. Z. Anorg. Allg. Chem. 2014, 640, 1127. (b) Lemouchi, C.; Yamamoto, H. M.; Kato, R.; Simonov, S.; Zorina, L.; Rodríguez-Fortea, A.; Canadell, E.; Wzietek, P.; Iliopoulos, K.; Gindre, D.; Chrysos, M.; Batail, P. Cryst. Growth Des. 2014, 14, 3375. (c) Comotti, A.; Bracco, S.; Yamamoto, A.; Beretta, M.; Hirukawa, T.; Tohnai, N.; Miyata, M.; Sozzani, P. J. Am. Chem. Soc. 2014, 136, 618. (d) Lemouchi, C.; Iliopoulos, K.; Zorina, L.; Simonov, S.; Wzietek, P.; Cauchy, T.; Rodríguez-Fortea, A.; Canadell, E.; Kaleta, J.; Michl, J.; Gindre, D.; Chrysos, M.; Batail, P. J. Am. Chem. Soc. 2013, 135, 9366.

(8) (a) Claridge, T. D. W. High Resolution NMR Techniques in Organic Chemistry; Pergamon: Oxford, U.K., 1999; Section 8.7.4. (b) Ernst, R. R.; Bodenhausen, G.; Wokaun, A. Principles of Nuclear Magnetic Resonance in One and Two Dimensions; Clarendon: Oxford, U.K., 1987. (c) Perrin, C. L.; Dwyer, T. J. Chem. Rev. 1990, 90, 935. (9) (a) Jeener, J.; Meier, B. H.; Bachmann, P.; Ernst, R. R. J. Chem. Phys. 1979, 71, 4546. (b) Gridnev, I. D.; Tok, O. L.; Gridneva, N. A.; Bubnov, Y. N.; Schreiner, P. R. J. Am. Chem. Soc. 1998, 120, 1034. (10) (a) Herndon, J. W. Coord. Chem. Rev. 2014, 272, 48. (b) Fürstner, A. Science 2013, 341, 1229713. (c) Grubbs, R. H., Ed. Handbook of Metathesis; Wiley-VCH: Weinheim, 2003; Vols. 1−3. (d) Schrock, R. R. Angew. Chem., Int. Ed. 2006, 45, 3748. (e) Grubbs, R. H. Angew. Chem., Int. Ed. 2006, 45, 3760. (f) Vougioukalakis, G. C.; Grubbs, R. H. Chem. Rev. 2010, 110, 1746.

(11) Cobas, J. C.; Martin-Pastor, M. EXSYCalc Ver 1.0; Mestrelab Research S.L.: A Coruñ a, Spain, 2007.

(12) (a) Maeda, H.; Maeda, T.; Mizuno, K. Molecules 2012, 17, 5108. (b) Shizuka, H.; Obuchi, H.; Ishikawa, M.; Kumada, M. J. Chem. Soc., Faraday Trans. 1 1984, 80, 383. (c) Ishikawa, M.; Oda, M.; Miyoshi, N.; Fábry, L.; Kumada, M.; Yamabe, T.; Akagi, K.; Fukui, K. J. Am. Chem. Soc. 1979, 101, 4612. (d) Pitt, C. G.; Carey, R. N.; Toren, E. C., Jr. J. Am. Chem. Soc. 1972, 94, 3806.

(13) Weber, G.; Teale, F. W. J. Trans. Faraday Soc. 1957, 53, 646.